

ABSTRACT

Thisisastudyconductedtoassesstheinterconnectionbetweenthemethanolicdraw out of *Lawsonia inermis* (leaf) and *Psidium guajava* (leaf) in combination withantimicrobial drugs including Ciprofloxacin and Amoxicillin against differentmicroorganismsi.e.

Staphylococcusaureus, Klebsiellapneumonia and Epidermophyton floccosum.

The well diffusion method is used to assess the interaction between methanolic extracts and antimic robial agents.

The experiments howed that methanolic extracts help increase the inhibition zones of Cipr of loxacin and Amoxicillin against the microbial strainstaken.

This study evaluates the inhibition zones and also concludes the synergisticactivityofantimicrobialdrugscombinedwithplantextractswhichmaysh owapossible wayto treat infections caused by *S.aureus*, *K.pneumonia* and *E.floccosum*.

KEYWORDS: Methanolextracts, synergism, ciprofloxacin, Amoxicillin, *Staphylococcusaureus*, *Klebsiellapneumonia*, *Epidermophytonfloccosum*.

INTRODUCTION

World's population sees deaths due to infections every single day. Infections arecaused by many known and unknown pathogens invading human systems which maycause symptoms, pain and ultimately death. Although several drugs have beendeveloped by pharmaceutical companies in past years, opposition to these drugs bybacteria has gone up over time which is now a major concern. ^[1] Microbialrefusal to accept to classical antibiotics and their fast increment has brought up seriousthinking in the cure of spreading diseases. Microbial spread are the reasonfor so many deaths each year worldwide. The happening of the advancement ofresistance is responsible for the available antibacterial drugs to becoming less powerful orevenineffective.Toget control

oftheresistanceofprescription, manystrategies have been suggested; one of them includes a mix of other molecules with imperfect antibiotics which restores the beneficial antibacterial activity.^[2]

Plantsarewidelyknownfortheirtherapeuticpropertiesandfromcenturiesofbeingused in the field of medicine can play a key role in eradicating such clinicalinfections. Plant extracts have been used in many experiments for such desirableresults as they contain many phytochemical compounds which may add strength inresistanceagainstthemicroorganisms.

Its fact, bacteria have the genetic power to pass on and have resistance to drug sused as the rapeutic representative. *S. aureus* (Gram-positive bacterium) and

K.pneumonia(Gram-negative bacterium) are recognized as the foremost reason of infections inpeople happening in both the community and hospitals. *S.aureus* and *K.pneumonia* are responsible for common respiratory infections while *E.floccosum* is a filamentousfungusthatisresponsible forskin-related infections.

Drugsynergismin the middle ofstudiedantimicrobialagents andbioactiveplantextractsis agreat concept that has been used widely now. ^[3] This study was carried out tosynthesize methanolic extract using *Lawsonia inermis* and *Psidium guajava* wereanalyzed for their antibacterial activity along with antimicrobial agents againstpathogenicbacteriain laboratoryconditions. Synergismisanewconceptindevelopingagentsforantimicrobialactivity. Thenew

approach is a therapy or mix of synergistic therapy in opposition to resistancemicroorganismswhichmake waytonewwaysofhandlinginfectiousdiseases.^[4] In present study, we reported the inhibition zones and synergistic work of themethanolic extracts of *Lawsonia inermis* (leaf) and *Psidium guajava* (leaf) in combinationwith antimicrobial drugs including Ciprofloxacin and Amoxicillin against differentmicroorganisms i.e. *Staphylococcus aureus*, *Klebsiella pneumonia* and *Epidermophytonfloccosum*.



Lawsoniainermis(L.)



Psidiumguajava(L.)

MethodsandMaterials

PLANTMATERIAL:

Driedleavesof*Lawsoniainermis*and*Psidiumguajava*are gatheredfromthe neighbourhoodof Visakhapatnam, Andhra Pradesh, India. The plant bits were rinsed and air-driedat roomtemperature.

PLANTEXTRACTPREPARATION:

The put together plant materials are cut into small pieces, shade dried andpowdered in a Willy mill. This material was put to weigh measure and extracted with Methanol along with a soxhlet extractor for 5-6 hours at a temperature not going beyondtheboilingpoint that the solvent

possess.Forevery100gramsofdrymaterial,2litresofsolventwere used. The taken out solvents were concentrated under reduced pressure with arotary evaporator. The residue received was considered as crude extracts and kept inafreezer untilassayed.1gofeachextract wastakenanddissolved in1mlofDiMethylSulphoxide (DMSO). Thus 500, 250 and 125mg/ml of stock were obtained as astandardconcentrationofextracts.

BACTERIALSTRAINS:

Threemothercultures of S. aureus,

K.pneumonia and *E.floccosum* obtained from Adhya Bioscience laboratories are used to eval uate the increase in inhibition zones and show the synergistic effect between plant extracts and antimicrobial drugs against the respective infections.

ANTIMICROBIALDRUGS:

Two drugs are used for the evaluation studies which include Ciprofloxacin andAmoxicillin.Desiredconcentrationsofantibioticdrugswerepreparedusingwaterasasolvent forciprofloxacinandamoxicillin solutions.

ANTIMICROBIALTESTS:

Well-diffusion method is used to measure the antibacterial activity in the experiment. Three Petri plates containing 20ml of Nutrient agar media were inoculated with a 24-hour culture of Bacterial strains. 4 wells of 6mm diameter each were punched in thePetri plates containing nutrient agar media. 3 wells in each plate were filled with125mg/ml, 250mg/ml and 500mg/ml standard concentrations of extracts/antibiotic

orcombinationrespectivelyand4thwellisfilledwith30µlofeitherwhichis consideredas a control. The Petri plates were incubated at 37°C for 24 hours. Assessment ofantibacterial activity is done by measuring the bacterial inhibition zones around thewell with a zone scale. The average of three replicates for each extract, antibiotic and combinationwasassayed.

ResultsandDiscussion

Different mechanisms of antimicrobial drugs are seen in this experiment.

Somenotable synergistic interactions (Amoxicillin with *L.inermis* and *P.guajava* against*S.aureus,K.pneumonia* and *E.floccosum*) and antagonistic interactions (Ciprofloxa cinwith *L.inermis* and *P.guajava* against *S.aureus* and *E.floccosum*) were identified.

While Ciproflox a cin showed no effect against

*K.pneumonia*whencombinedwithplantextracts.Thedatarepresentedbelowshowthepotenti aleffectofantimicrobialdrugs incombination with plant extracts used against *S.aureus, K.pneumonia* and *E.floccosum*.[Table1,2,3]

| DRUG TARGET | DRUG | Psidium guajava | Lawsonia inermis | SYNERGISM RATE[EXTRACT/ DRUG] | MEAN |
|---|---------------|--------------------|---------------------|-------------------------------------|------|
| Nucleic acidsynthe sisinhibito r | Ciprofloxacin | А | А | 0 | 0 |
| CellWall Biosynthesis inhibitor | Amoxicillin | S | S | 2 | 2 |
| TOTAL 2 | | 1 | 1 | - | - |

TABLE1-INHIBITION EFFECTAGAINST*Staphylococcusaureus*

(A) –Antagonism;(S)–Synergism;(Nochange)

TABLE2-INHIBITIONEFFECTAGAINSTKlebsiellapneumonia

| DRUG TARGET | DRUG | Psidium guajava | Lawsonia inermis | SYNERGISM RATE[EXTRACT/ DRUG] | MEAN |
|---|---------------|--------------------|---------------------|-------------------------------------|------|
| Nucleic acidsynthe sisinhibito r | Ciprofloxacin | Nochange | Nochange | - | - |
| CellWallBio synthesisinh ibitor | Amoxicillin | S | S | 2 | 2 |
| TOTAL | 2 | 1 | 1 | - | - |

(A) -Antagonism;(S)-Synergism;(Nochange)

| DRUG TARGET | DRUG | Psidium guajava | Lawsonia inermis | SYNERGISM RATE[EXTRACT/ DRUG] | MEAN |
|---------------------------------------|---------------|--------------------|---------------------|-------------------------------------|------|
| Nucleicacid synthesisinhi bitor | Ciprofloxacin | А | А | 0 | 0 |
| CellWallBio synthesisinh ibitor | Amoxicillin | S | S | 2 | 2 |
| TOTAL 2 | | 1 | 1 | - | - |

TABLE3-INHIBITIONEFFECTAGAINSTEpidermophytonfloccosum

(A)–Antagonism; (S)–Synergism; (Nochange)

Respective inhibition zones assayed in this experiment are mentioned following the plantextractstaken[Table4, 5, 6].AssayofInhibitionis donebytakingzeroasnegativecontroland positive control for a respective antibiotic used against the respective microorganismvaried.

TABLE4– INHIBITION ZONES OF P. guajava

| <u>Psidiumguajava</u> (Leaf) | | | | | | | | |
|------------------------------|---|--------------------|--------------------|---------------------|---------------------------------------|--------------------|--------------------|--------------------------|
| MICROBIAL | Ciprofloxacin + Plant extract(mg/ml) | | | Ciprofloxacin | Amoxicillin + Plant extract(mg/ml) | | | |
| STRAIN | 125mg/ml (30µl) | 250mg/ml (30µl) | 500mg/ml (30µl) | (control)(30 µl) | 125mg/ml (30µl) | 250mg/ml (30µl) | 500mg/ml (30µl) | Amoxicillin (control) |
| | | | | | | | | (30µl) |
| S.aureus | 19mm | 20mm | 20mm | 28mm | 15mm | 15mm | 16mm | 11mm |
| K.pneumonia | 37mm | 37mm | 40mm | 40mm | 18mm | 20mm | 21mm | 16mm |
| E.floccosum | 33mm | 31mm | 28mm | 40mm | 16mm | 18mm | 22mm | 19mm |

| Lawsoniainermis (Leaf) | | | | | | | | |
|------------------------|---|--------------------|--------------------|---------------------|---------------------------------------|--------------------|--------------------|--------------------------|
| | Ciprofloxacin + Plant extract(mg/ml) | | | Ciprofloxacin | Amoxicillin + Plant extract(mg/ml) | | | |
| MICROBIAL STRAIN | 125mg/ml (30µl) | 250mg/ml (30µl) | 500mg/ml (30µl) | (control)(30 µl) | 125mg/ml (30µl) | 250mg/ml (30µl) | 500mg/ml (30µl) | Amoxicillin (control) |
| | | | | | | | | (30µl) |
| S.aureus | 21mm | 23mm | 26mm | 28mm | 10mm | 11mm | 11mm | 11mm |
| K.pneumonia | 35mm | 39mm | 40mm | 40mm | 17mm | 20mm | 24mm | 18mm |
| E.floccosum | 30mm | 31mm | 33mm | 40mm | 16mm | 17mm | 22mm | 21mm |

TABLE5-INHIBITIONZONESOFL.inermis

Itisrecognizedthat*S.aureus*isoneoftheleadingcausesofinfectionsthatoccurinboththe community and the hospital. As with many nosocomial pathogens, Multidrug-resistant Staphylococci are extremely difficult to treat because they are resistant toalmost all antibiotics clinically available right now and also can cause meningitis. ^[5] Anew approach to solving the bacterial resistance problem depends on the ability of plantextracts to act synergistically with antibiotics. ^[5] In this study, reference andenvironmental strains of pathogenic organisms were used to examine drug resistance inclinical settings often associated with these organisms. To determine the effects ofcombinationswithantibiotics, extracts ofplantsareused.^[6]

Thetherapeutic potentialofleafextractsfrom*L.inermis*and*P.guajava*hasdemonstrated the strong synergy between crude methanolic extract of the leaves andfirst-line antibiotics resulting in clinically useful applications against respectivemicrobialstrains.^[5,6]Bacterialinfectionscanbetreatedwithadvantageoussynergistic effectswhencombinedantibiotictherapy isused. This novel concept of synergism can either be beneficial (additive/synergisticinteractions) or deleterious (Antagonistic/toxic interactions).^[7] A study reported

asynergisticeffectbetweentheplantextractsandthecellwallbiosynthesisinhibitoragainst*S* .*aureus* and*K.pneumonia*while notmuchis knownabout*E.floccosum*.

In previous studies, amoxicillin was secured to synergize well with the variousphytochemicalcompounds.However,Nucleicacidsynthesisinhibitorsandplantex tractshadnosynergisticactivity.^[5]

In gram-positive and gram-negative pathogens such as *Staphylococcus aureus* and *Klebsiella pneumonia*, MDP pumps, which recognized and remove a variety ofcompounds with unrelated structures from bacterial cells, have been identified. In vitro, it has been demonstrated that some compounds can work synergistically with antibiotics to modify the resistance phenotype in bacteria. ^[8] As they exhibit a low risk of causing bacterial resistance to their actions, combinations of antibiotics and plant extracts are likely to provide the basis force reating new approaches in resistance modifying agents.

These extracts have a combination of different bioactive compounds, which makesmicrobialadaptionextremelydifficultcomparedtosinglecomponentantibiotics.^[5]

Our research indicates that plant extracts strengthen theantimicrobialeffectswhencombinedwithdrugsagainstclinicalinfectionsandcanreduc ethespreadofbacteria.^[7]

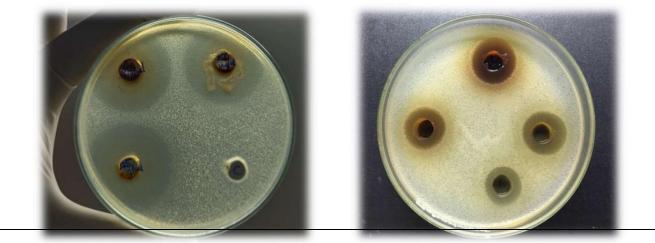


Fig:Acloserviewofinhibitionzonesobservedintheexperiment.

Conclusion

This study concluded that the extracts of *L.inermis* and *P.guajava* are found to have the capacity of increasing the susceptibility of the studied microbial strains to various antimicrobial drugs. The present study clearly states the possibility of the use of the above shown synergistic drug (amoxicillin)-

plantcombinationsforcombatinginfectious diseases caused by *S.aureus*, *K.pneumonia* and *E.floccosum* whereas

ciproflox a cin showed an egative effect when combined with both plant extracts.

The results represented in this respective report were encouraging in correlation withamoxicillin,althoughclinicalcontrolledstudies

areneededtodefinetherealefficacyand possible toxic effects in vivo. This study majorly suggests the possibility of concurrent use of these antimicrobial drugs and extracts in combination in treating infections caused by respective strains used and plants may play a key future part inclinical studies.

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